

# LOW COST, ULTRA-HIGH THROUHPUT PARTICLE COUNTING USING INERTIAL MICROFLUIDICS

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## ABSTRACT

In this work, an ultra-high throughput microfluidic particle counting system is demonstrated. For the particle counting, a low cost custom-design optical hardware is developed. The microfluidic chip utilizes the inertial microfluidics to focus the particles in a certain location which significantly enhanced the optical signal utilized for the quantification of the number concentration. The effect of the particle focusing on the counting performance is demonstrated. The proposed system has a potential to be portable and has a capability to process 10 ml of sample within couple minutes.

**KEYWORDS:** Inertial microfluidics, Particle counting, Fluorescence microscopy

## INTRODUCTION

Particle counting is one of the important and key processes regarding the biotechnological and biomedical applications<sup>1</sup>. Although an accurate enumeration of target bio-particles (with low number concentration) is desired in many applications, there are also many application in which the counting of high number of particles is desired such as red blood cell counting, somatic cell and bacterial counting in whole milk<sup>2</sup>.

## THEORY

Typically, the Reynolds number in conventional microfluidic applications is in the order of  $10^{-2}$ -1.0 which dictates Stoke's flow and streamlines parallel to the channel walls. However, when the Reynolds number reaches 10-50, some secondary flows begin to occur within the channel which induces lift drag and forces on the particles flowing in the channel. The balance of these forces dictates motion of particles at some certain locations (both in the lateral and height direction)<sup>3</sup>. These applications are known as inertial microfluidics. This focusing effect is more pronounced if the flow is within a curved channel. In this work, this focusing nature of the inertial microfluidics is utilized to amplify the optical signal for particle counting.

## EXPERIMENTAL

The fabrication of the molds are performed by high-precision machining. Two channels, one is straight (H:200µm, W:750µm) and one is spiral (H:600µm, W:70µm) with 5 runs are fabricated. The experimental setup consists of a laboratory syringe pump (New Era Pump Systems-NE 300), syringes and tubing. A buffer solution in which 10µm fluorescent (Distrlab Fluoro-Max) particles are suspended in a deionized (DI) water solution. Particle solution with different concentrations are used in the experiments. The whole system together with the custom-design optical system are shown in Figure 1. As delineated in Figure 1-(B), a custom-made camera-based microfluidic cytometer is built for optical determination of particle concentration in flow. The fluorescent counting system employs a 4X (0.1 NA) objective lens to view the microchannel from below. Excitation, emission and dichroic filters are selected based on the spectral characteristics of green fluorescent particles (excitation and emission peak at 468 nm and 508 nm, respectively). A narrow band LED is used as a source for excitation. A low-cost CMOS board camera (Imaging source GmbH) is utilized to measure fluorescent intensity captures images at every 1 s with 0.25 s exposure time, providing sufficient enough intensity for measurements at low particle concentrations. The optical magnification is adjusted to visualize the entire channel width with the projected pixel size of 1.15 µm on the camera sensor. A motorized stage (Psaron HTI) is integrated into the system to perform axial-scan over the channel height with 1.3 µm resolution. Image data from the axial-scan is used to find the hydrodynamic depth of focus.

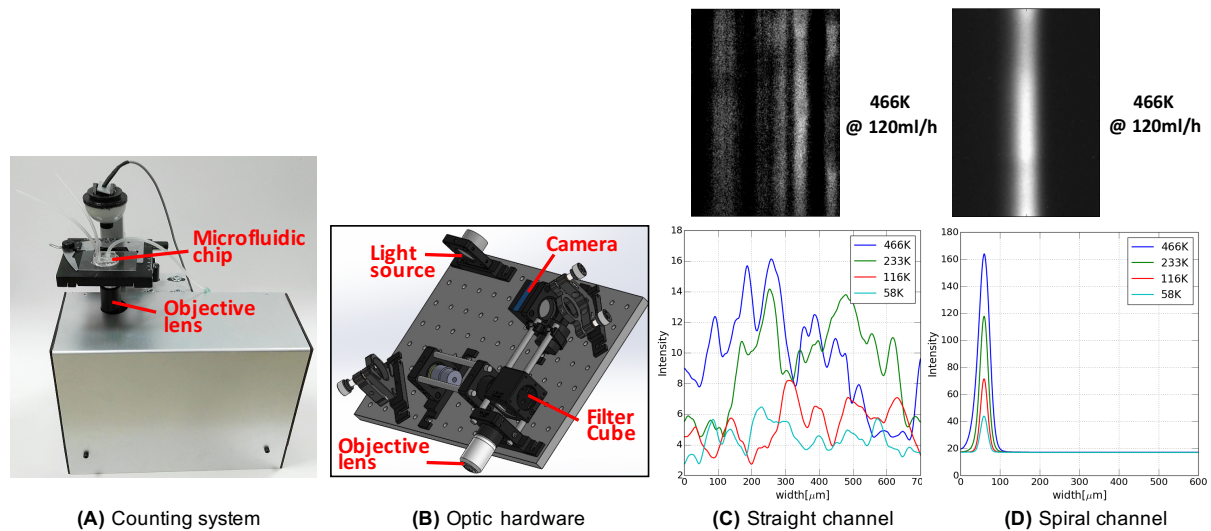


Figure 1: Hardware of the system and the experimental results.

## RESULTS AND DISCUSSION

The images acquired at a fixed flow rate of 120 ml/h are first averaged over the flow direction in order to obtain the fluorescent intensity as a function of distance over the channel width. The acquisition time is 20 s during which 670  $\mu$ l volume of fluid is analyzed. During this short period of time, as shown in Figure 1-(D), the intensity profile of focused particles on the spiral channel is greatly influenced by the particle concentration. As depicted from the figure, as the concentration increases, the change in the peak intensity per thousand particles is between 0.12 and 0.67%. On the other hand, on the straight channel without the focusing effect the intensity profiles for different number of particles are indistinguishable from each other. The intensity profile also shows the focusing effect of the spiral channel.

## CONCLUSION

The determination of the particle concentration is obtained for 10 $\mu$ m successfully with ultra high-throughput. The extension of this study will be the generating experimental results at different flow rates together with 2 $\mu$ m particles. 2 $\mu$ m particles represents the bacteria and 10 $\mu$ m particles resembles the somatic cells. Therefore, ultimate goal of this project is to utilize this system to determine the bacterial load and somatic cell count of the whole milk with a counting precision and accuracy less than 3% and 10%, respectively. These values represents the capabilities of the counters currently available in the market, measuring only up to 10-20  $\mu$ l of the sample one at a time.

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